



# Toward the standardization of mitochondrial proteomics

*Luisa Pieroni*

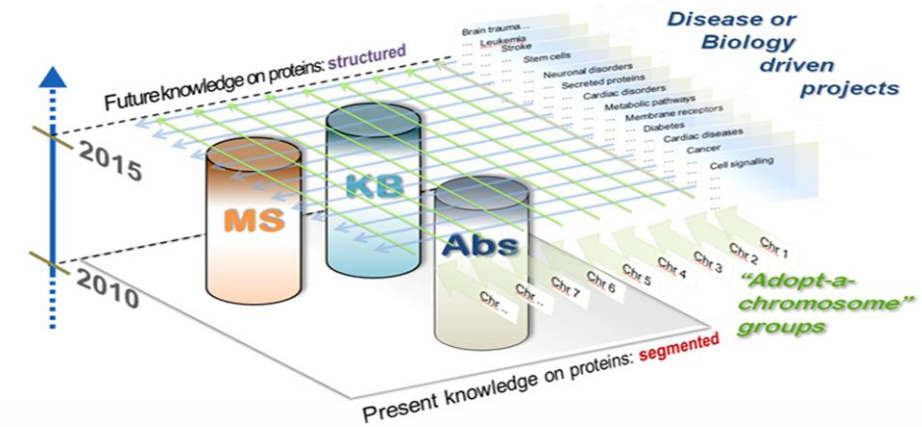
*Proteomics and Metabonomics*

*IRCCS Fondazione S.Lucia-Rome*

*Italy*

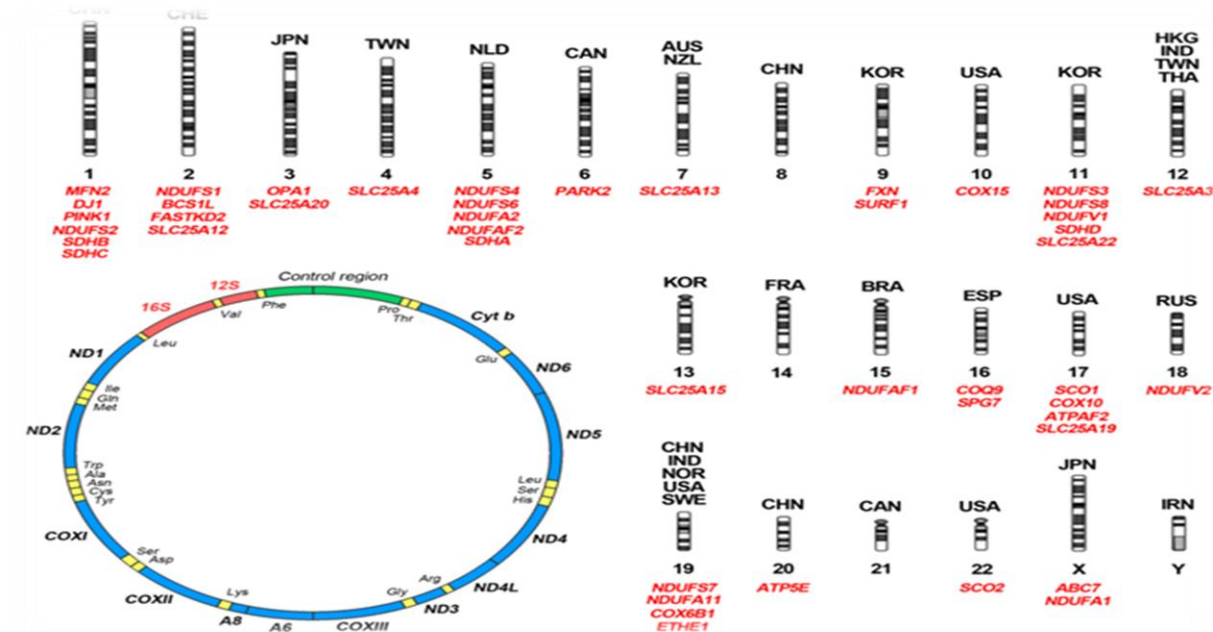
**MiP 2017/MitoEAGLE Conference, Hradec Králové**  
**November 17, 2017**

Mapping of the entire human proteome using currently available and emerging techniques



## C-HPP: Chromosome-Centric

## B/D-HPP: Biology/Disease



# Italian initiative of the Human Proteome Project dedicated to mitochondria

Journal of  
**proteome**  
research

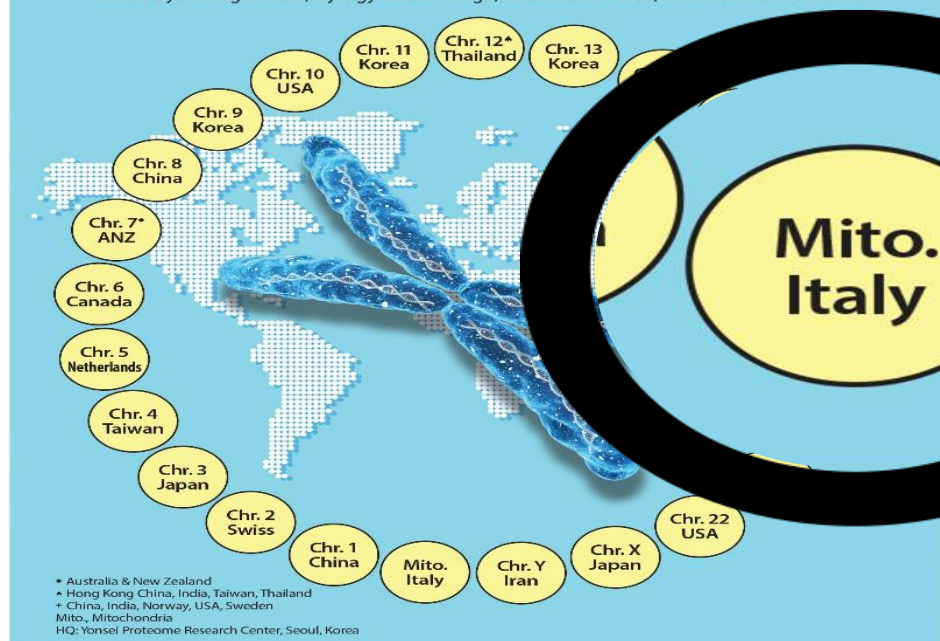
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## Chromosome-Centric Human Proteome Project

www.c-hpp.org 2012 - 2022

Edited by: Young-Ki Paik, Gyorgy Marko-Varga, Gilbert S. Omenn, and William S. Hancock

Molecular  
BioSystems

RSC Publishing

REVIEW

View Article Online  
View Journal | View IssueCite this: *Mol. BioSyst.*, 2013,  
9, 1984

## The mitochondrial Italian Human Proteome Project initiative (mt-HPP)<sup>†</sup>

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Mitochondria carry maternally inherited genetic material, called the mitochondrial genome (mtDNA), which can be defined as the 25th human chromosome. The chromosome-centric Human Proteome Project (c-HPP) has initially focused its activities addressing the characterization and quantification of the nuclear encoded proteins. Following the last International HUPO Congress in Boston (September 2012) it was clear that however small the mitochondrial chromosome is, it plays an important role in many biological and physiopathological functions. Mutations in the mtDNA have been shown to be associated with dozens of unexplained disorders and the information contained in the mtDNA should be of major relevance to the understanding of many human diseases. Within this paper we describe the Italian initiative of the Human Proteome Project dedicated to mitochondria as part of both programs:

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www.rsc.org/molecularbiosystems

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## Mitochondrial proteomics investigation of a cellular model of impaired dopamine homeostasis, an early step in Parkinson's disease pathogenesis†

Tiziana Alberio,<sup>ab</sup> Heather Bondi,<sup>ab</sup> Flavia Colom,<sup>de</sup> Luisa Pieroni,<sup>de</sup> Andrea Urbani<sup>\*de</sup> and Mauro Fasella<sup>†</sup>

Molecular  
BioSystems



PAPER



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Citation: *Transl Psychiatry* (2016) 6, e904; doi:10.1038/tp.2016.194  
[www.nature.com/tp](http://www.nature.com/tp)

### ORIGINAL ARTICLE

Bottom-up proteomics suggests an association between differential expression of mitochondrial proteins and chronic fatigue syndrome

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## Glucagon-like peptide 1 protects INS-1E mitochondria against palmitate-mediated beta-cell dysfunction: a proteomic study†

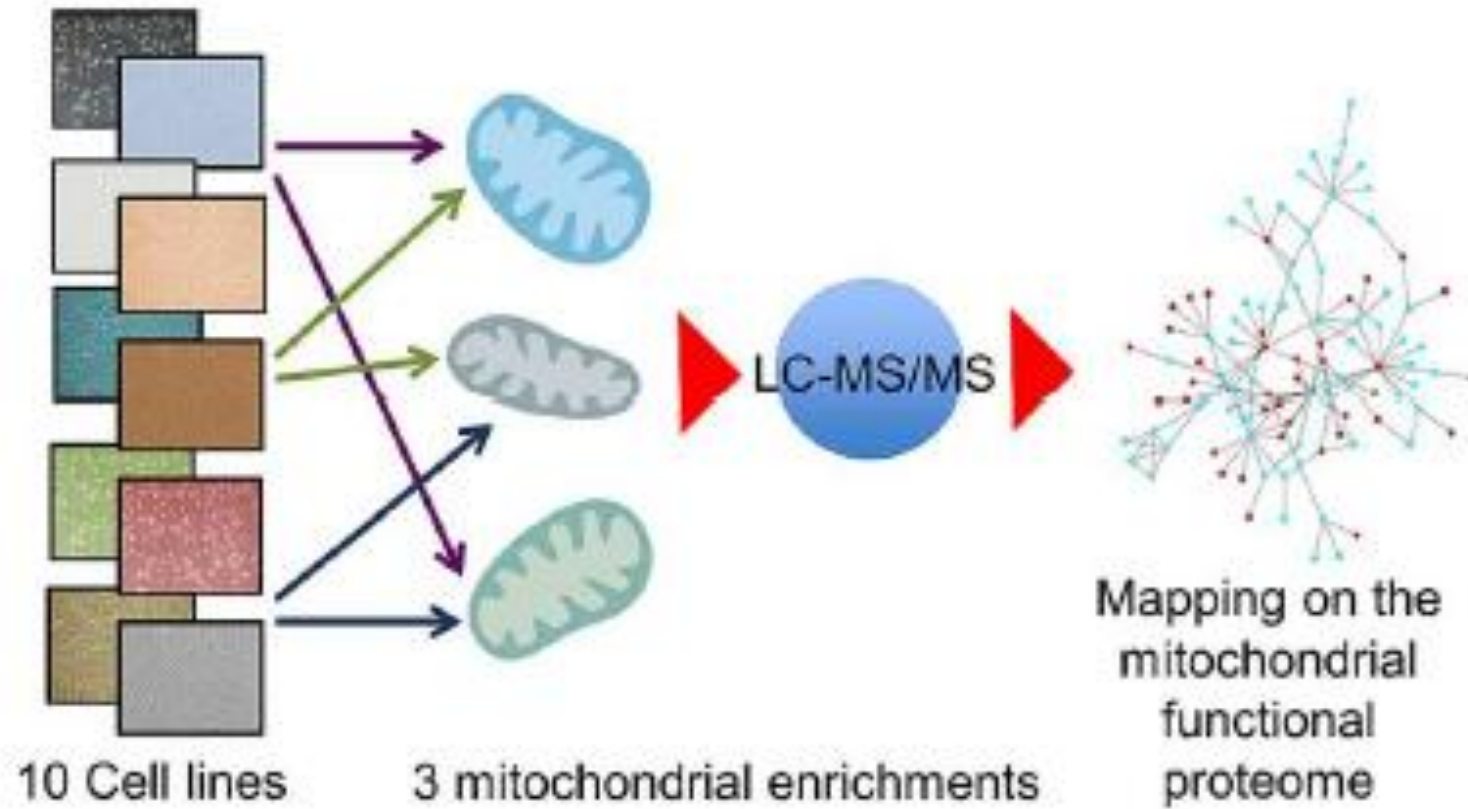
Federica Ciregia,<sup>ab</sup> Laura Giusti,<sup>a</sup> Maurizio Ronci,<sup>bcd</sup> Marco Bugliani,<sup>e</sup> Isabella Piga,<sup>a</sup> Luisa Pieroni,<sup>b</sup> Claudia Rossi,<sup>c</sup> Piero Marchetti,<sup>e</sup> Andrea Urbani<sup>bf</sup> and Antonio Lucacchini<sup>\*a</sup>



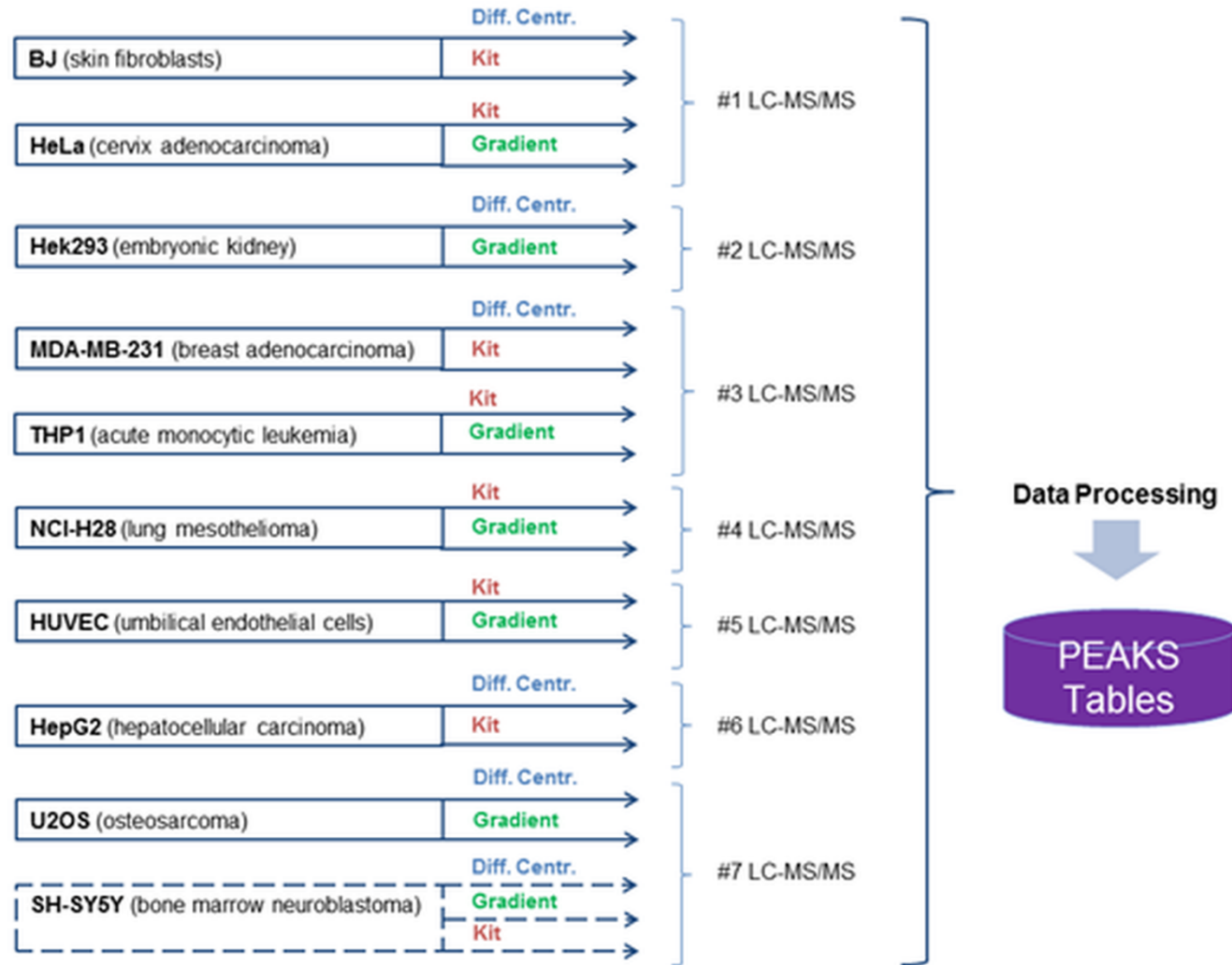
# We needed a mt-enrichment procedure standardization

- To address the need of our proteomics community to analyse complete mt-proteomes ( including Matrix, IMM, OMM and IMS proteins)
- To perform reliable comparison of data from different laboratory

# The study design



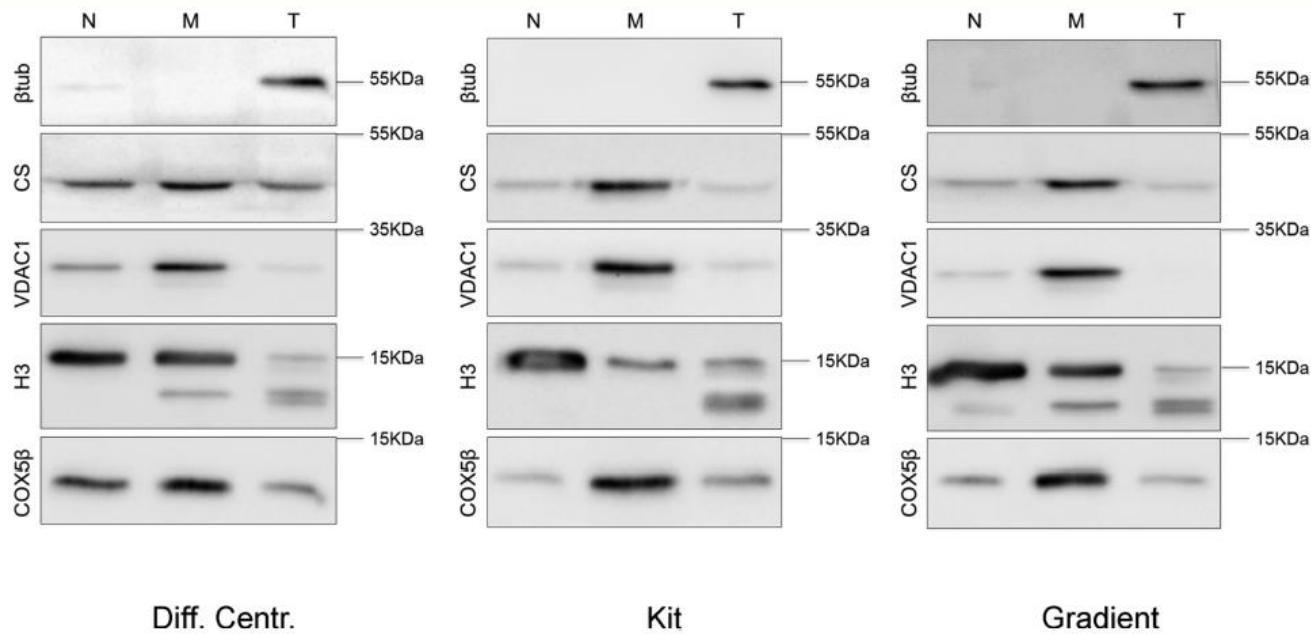
# The study design



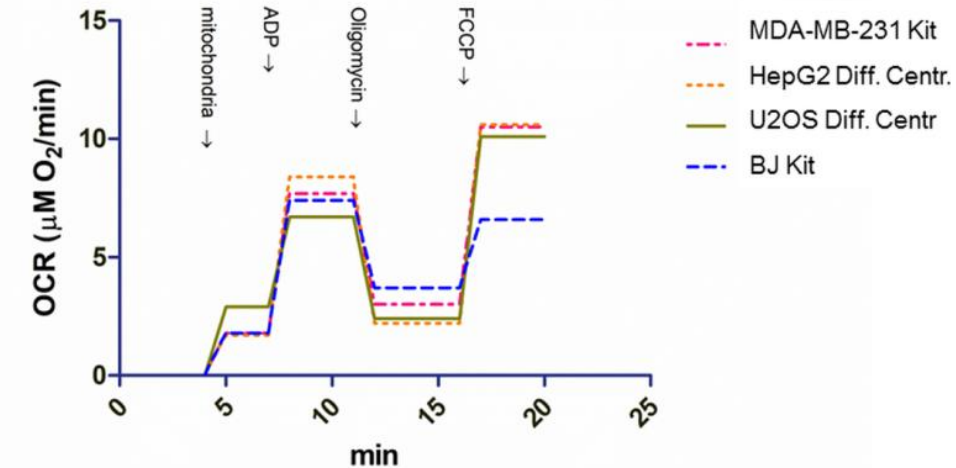
**Table 1. MS and Chromatographic Platforms**

no.	LC system	column	mass spectrometer	cells
1	nanoAcquity M Class	Waters HSST3 C <sub>18</sub> 75 $\mu$ m $\times$ 15 cm	Synapt G2 Si	BJ, HeLa
2	Dionex Ultimate 3000	Thermo Easy-Spray PepMap RSLC C <sub>18</sub> 75 $\mu$ m $\times$ 50 cm	Orbitrap Fusion	Hek293
3	Dionex Ultimate 3000	MS Wil GmbH C <sub>18</sub> 75 $\mu$ m $\times$ 20 cm	LTQ-Orbitrap- Velos	MDA-MB231, THP1
4	nanoEASY II	Nanoseparations C <sub>18</sub> 100 $\mu$ m $\times$ 20 cm	LTQ-Orbitrap-XL	NCI-H28
5	Dionex Ultimate 3000	Thermo Easy-Spray PepMap RSLC C <sub>18</sub> 75 $\mu$ m $\times$ 50 cm	Bruker Impact HD	HUVEC
6	Ekspert nanoLC 400	Thermo Acclaim PepMap 100 75 $\mu$ m $\times$ 25 cm	TripleTOF 5600+	HepG2
7	nanoEASY II	Thermo Acclaim PepMap 100 75 $\mu$ m $\times$ 25 cm	Bruker maXis HD	SH-SY5Y, U2OS

# Mt-enrichment yield and purity check



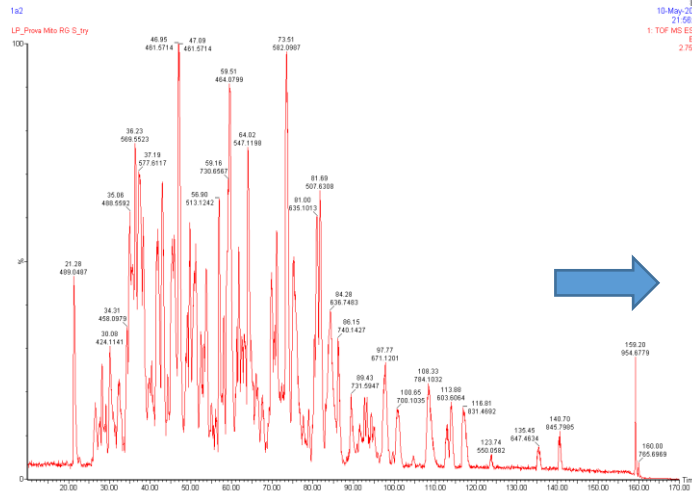
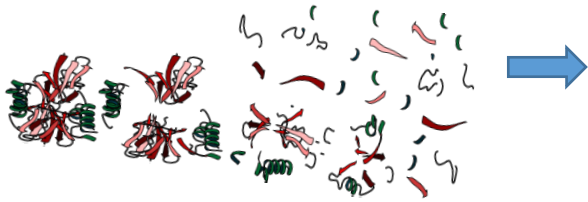
Cell line/isolation protocol	nmol/min mg prot before lysis	nmol/min mg prot after lysis	% ruptured
HepG2 Diff. Centr.	18.2	253.7	7.2
MDA-MB-231 Kit	17.2	137.1	12.3
HeLa Kit	30	180	16.7
SH-SY5Y Diff. Centr.	17.3	219	7.9
BJ Kit	26.8	237	11.3





# Label free shotgun “bottom up” proteomics

Proteins Tryptic digestion



LC-MS

Fusion OrbiTrap



maXis HD QTOF



Synapt G2si QTOF  
Waters

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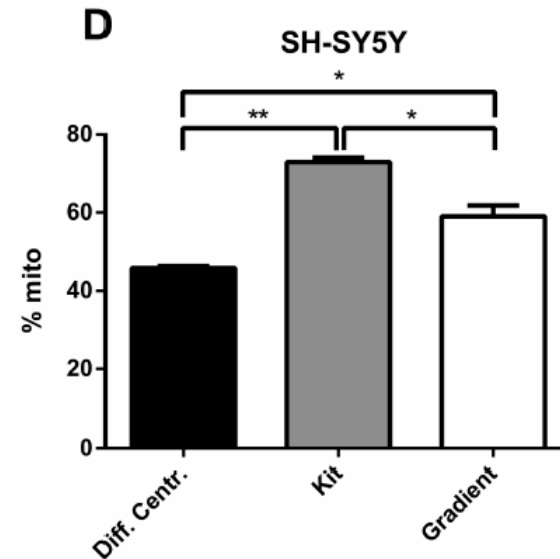
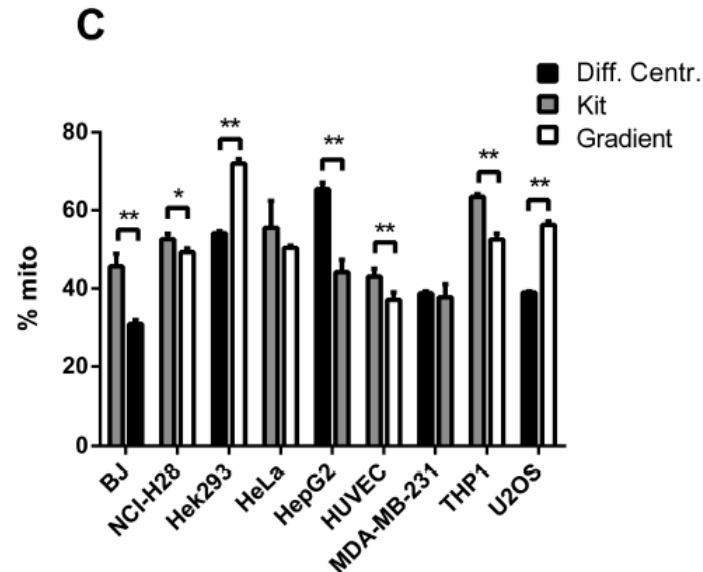
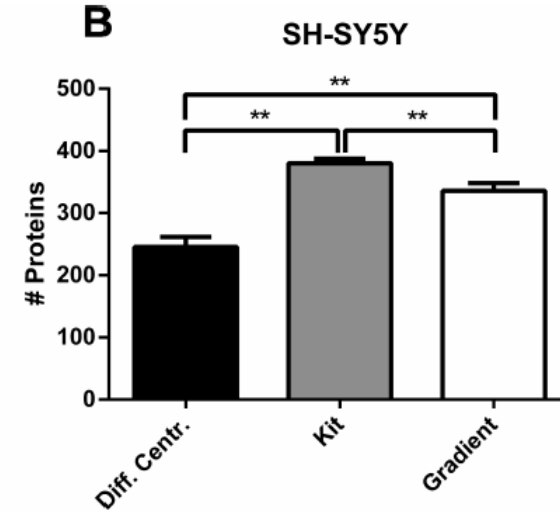
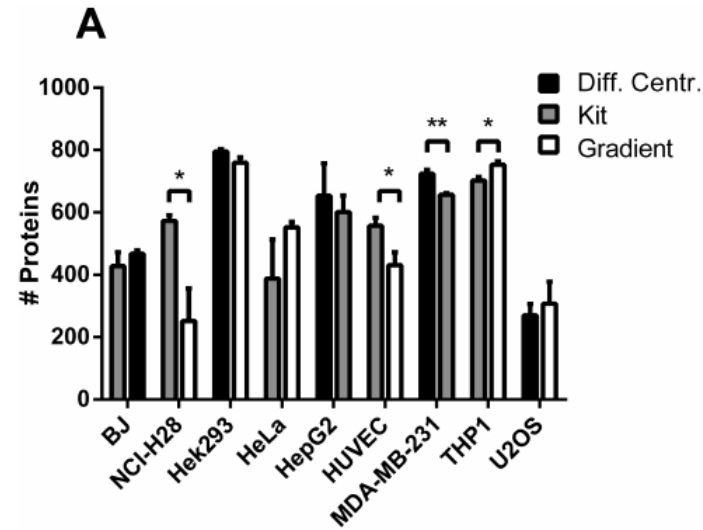


TripleTOF™ 5600



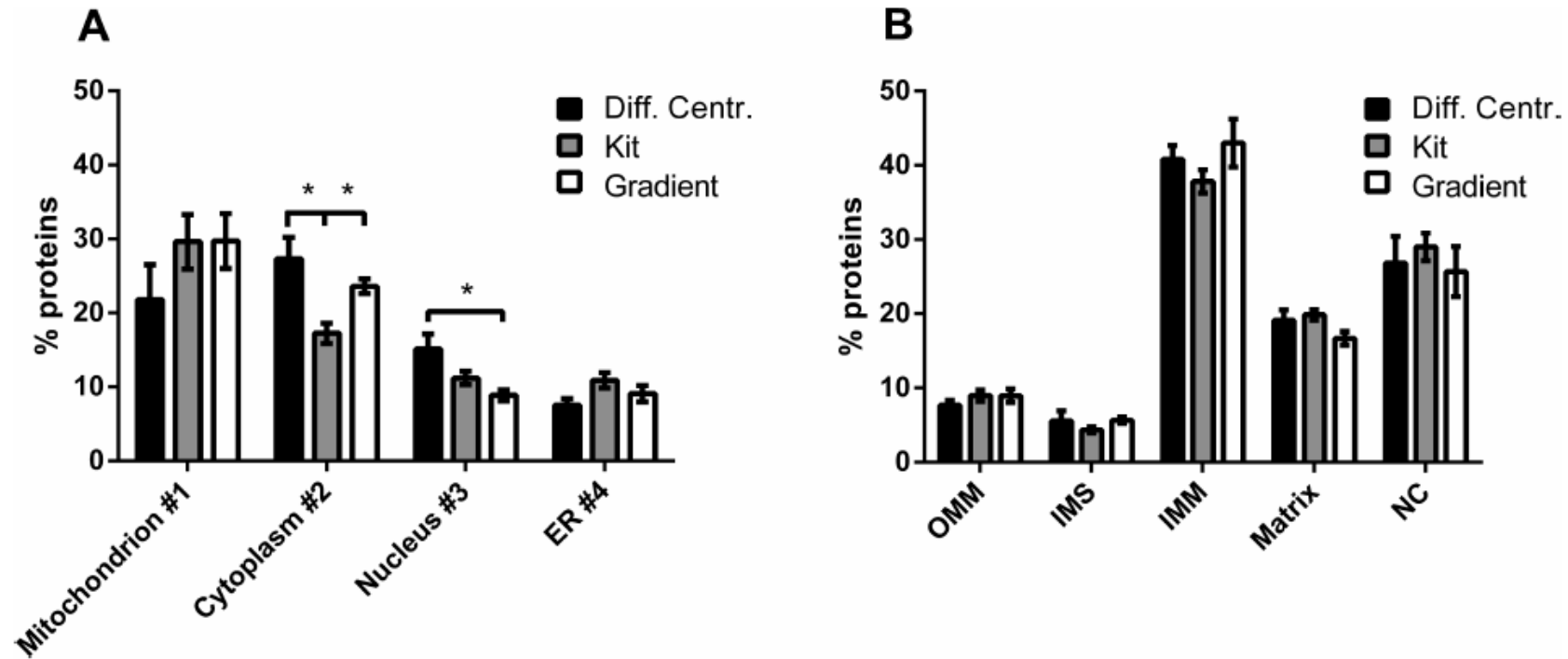


# Total and mt proteins <sup>(1)</sup> per cell line



<sup>(1)</sup> MitoCarta & IMPI

# Top subcellular locations and submitochondrial enrichment (2)



(2) Primary location in UniProt annotation

# Mapping on the functional mt-proteome

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EuPA Open Proteomics

journal homepage: [www.elsevier.com/locate/euprot](http://www.elsevier.com/locate/euprot)



ELSEVIER



Towards a functional definition of the mitochondrial human proteome

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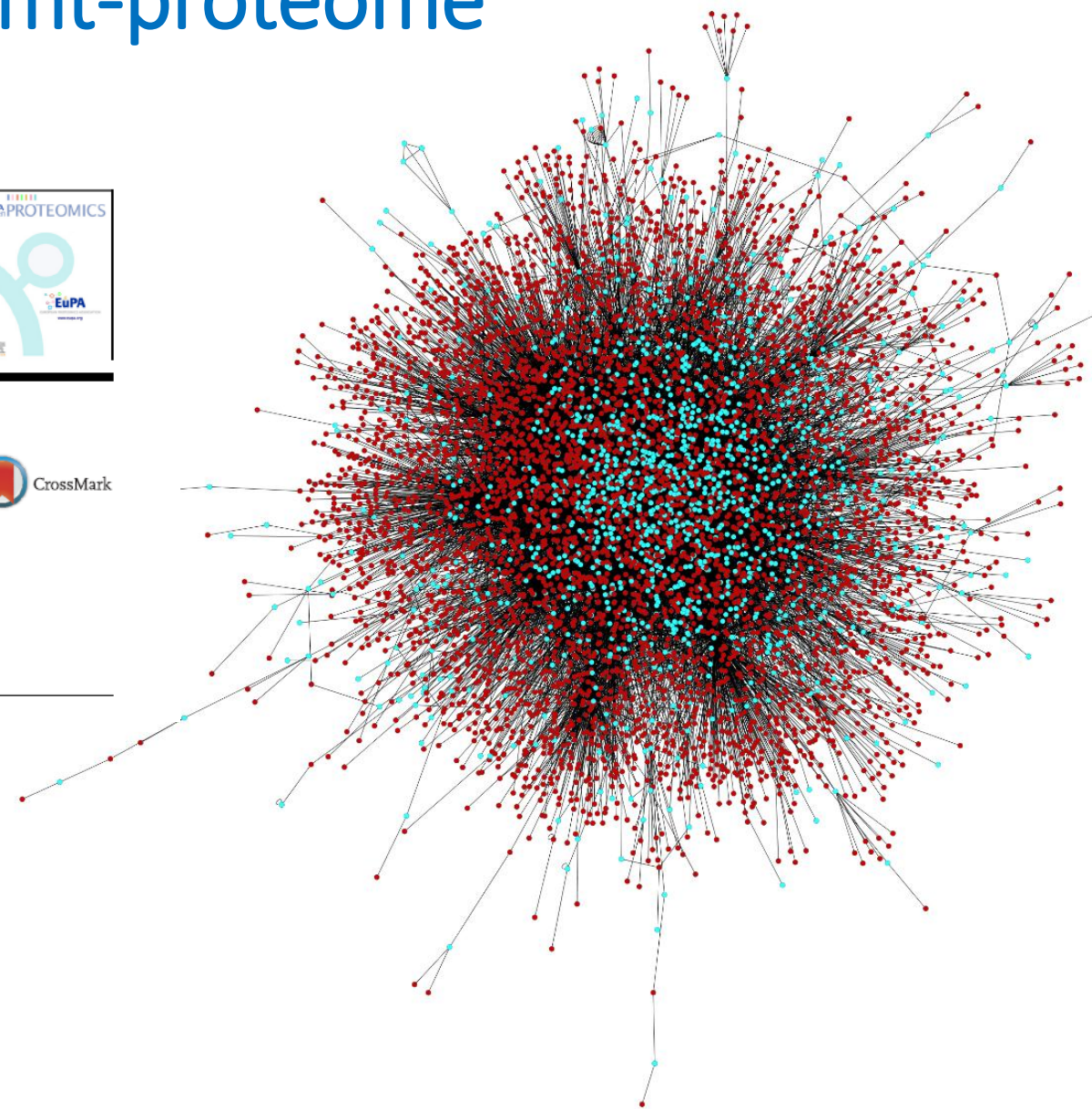
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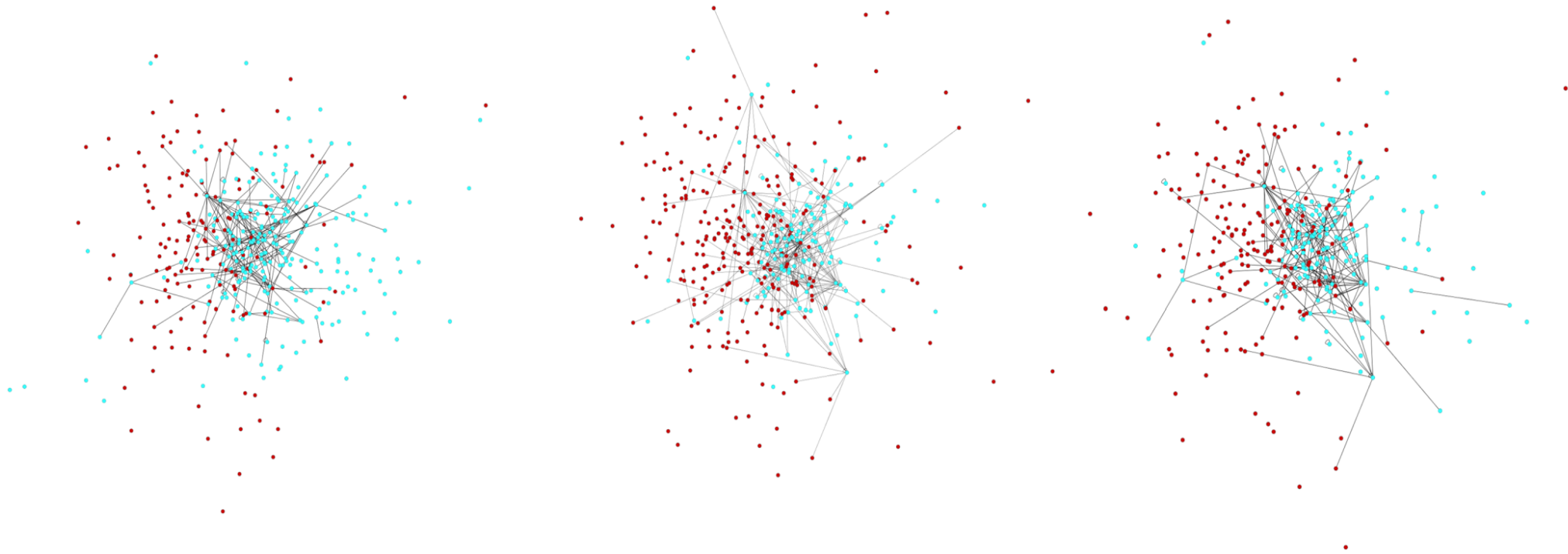
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# Mapping on the functional mt-proteome



Diff. Centr.

Kit

Gradient

# Mapping on the functional mt-proteome

cell line	method	% mapped	% mitochondrial	% clustered	% mitoclustered
BJ	kit	77	36	22	53
	diff. centr.	84	21	30	43
NCI-H28	kit	84	46	29	56
	gradient	88	38	28	65
Hek293	diff. centr.	87	44	41	58
	gradient	84	63	38	70
HeLa	kit	82	46	29	70
	gradient	86	41	34	56
HepG2	diff. centr.	81	57	32	69
	kit	78	38	25	65
HUVEC	kit	79	38	33	56
	gradient	82	30	37	49
MDA-MB-231	diff. centr.	79	32	38	49
	kit	76	36	36	53
THP1	kit	76	63	36	72
	gradient	78	51	38	62
U2OS	diff. centr.	83	30	29	53
	gradient	87	45	30	66
SH-SY5Y	diff. centr.	89	33	28	54
	kit	90	60	26	66
	gradient	89	47	32	64

# We do not draw unique conclusions but:

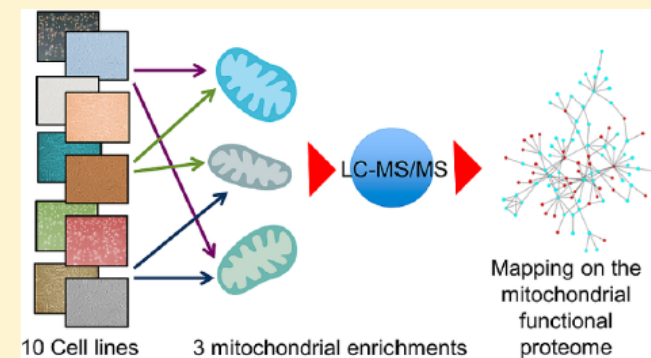
- We compared suitable procedures to achieve effective MS analysis of mitochondrial proteome, defining guidelines for different experimental designs, irrespectively of MS technological platforms available
- Standardization action will contribute to mt-HPP and B/D-HPP
- We added several mitochondrial datasets to specific proteomics KB (e.g. Proteome Xchange) as reference for future proteomics studies
- We integrated proteomic results from the functional point of view

# **Toward the Standardization of Mitochondrial Proteomics: The Italian Mitochondrial Human Proteome Project Initiative**

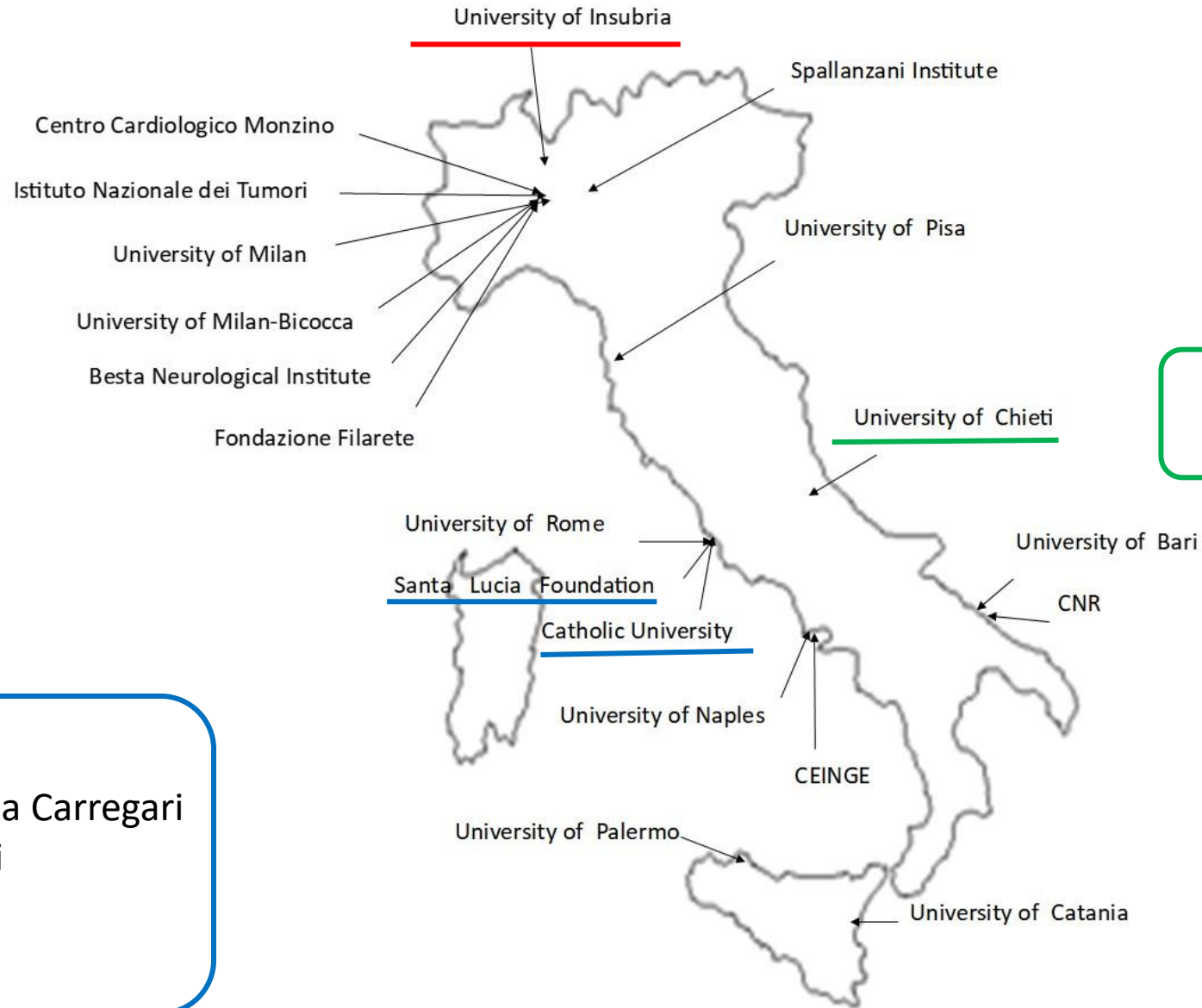
Tiziana Alberio,<sup>†,Φ</sup> Luisa Pieroni,<sup>‡,Φ</sup> Maurizio Ronci,<sup>‡,§,Φ</sup> Cristina Banfi,<sup>⊥</sup> Italia Bongarzone,<sup>¶</sup>  
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Rosaria Saletti,<sup>■</sup> Roberto Scatena,<sup>#</sup> Alessio Soggiu,<sup>●</sup> Gabriella Tedeschi,<sup>●,△</sup> Mara Zilocchi,<sup>†</sup>  
Paola Roncada,<sup>○</sup> Andrea Urbani,<sup>‡,#</sup> and Mauro Fasano<sup>\*,†,ID</sup>

**ABSTRACT:** The Mitochondrial Human Proteome Project aims at understanding the function of the mitochondrial proteome and its crosstalk with the proteome of other organelles. Being able to choose a suitable and validated enrichment protocol of functional mitochondria, based on the specific needs of the downstream proteomics analysis, would greatly help the researchers in the field. Mitochondrial fractions from ten model cell lines were prepared using three enrichment protocols and analyzed on seven different LC–MS/MS platforms. All data were processed using neXtProt as reference database. The data are available for the Human Proteome Project purposes through the ProteomeXchange Consortium with the identifier PXD007053. The processed data sets were analyzed using a suite of R routines to perform a statistical analysis and to retrieve subcellular and submitochondrial localizations. Although the overall

*Continued...*



# THANKS to the CONSORTIUM



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Federica Marini

Andrea Urbani